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Research Article

New records of the Ethiopian long-eared bat *Plecotus* balensis in central Ethiopia

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Abstract

The diversity of bats in Ethiopia comprises at least 80 species, among them the Ethiopian longeared bat that was described in 2000. It is most likely endemic to the highlands of Ethiopia. However, knowledge of the distribution of the species is limited. During a bat survey in 12 regions of central Ethiopia stretched over 700 km along the Ethiopian Rift, we trapped longeared bats at sites in three regions and confirmed the species' identity by molecular analysis. All occurrence sites of *P. balensis* were above 2500 m, confirming this taxon as a high-altitude species. Two of the regions are additions to the known range of *P. balensis* but it is most likely present in more high-altitude areas of Ethiopia than currently known. Additional surveys in so far unsampled areas are therefore indicated.

Keywords: Chiroptera, Vespertilioninae, Horn of Africa, distribution range, molecular species identification, cytochrome b

Introduction

Long-eared bats (genus *Plecotus*, Vespertilioninae) are primarily species of the temperate zone of Eurasia but also occur around the Mediterranean Sea and in the Nile Valley up to northern Sudan (Benda et al., 2004; Juste et al., 2004). The internal taxonomy of the genus has changed substantially in recent years due to the application of molecular analyses for species identification (Kruskop et al., 2020). Several species have only recently been described (Mucedda et al., 2002; Benda et al., 2004; Spitzenberger et al., 2006; Dolch et al., 2021). One of these is the Ethiopian long-eared bat *Plecotus balensis* Kruskop & Lavrenchenko, 2000. It is the most southern member of the genus and the only Afrotropical species (Benda et al., 2004). It was first described from the Harenna Forest in the Bale Mountains National Park (Kruskop & Lavrenchenko, 2000). Subsequently, it was confirmed at additional sites in the Ethiopian Highlands including the Simien Mountains, Mount Abune Yosef, the Guassa Community Conservation Area, Chilallo Mountains, and several more sites in the Bale Mountains (Juste et al., 2004; Razgour et al., 2019). It is believed to be endemic to the Ethiopian Highlands, but its exact distribution is unknown so far.

The *Plecotus* species of Ethiopia (Rüppell, 1842, Shewa) and Eritrea (Sordelli, 1902; Senna, 1905; Asmara) have been first recognized as *P. auritus*, but later renamed as *P. austriacus*, most likely belonging to the subspecies *christii* Gray, 1838 (Largen et al., 1974). This subspecies was later elevated to full species *P. christii*. Yalden et al. (1996; specimen in the World Museum, Liverpool) reported *P. austriacus* from two sites in the Bale Mountains which most likely are *P. balensis* (Juste et al., 2004). Whether the specimen from Eritrea belongs to *P. balensis* or *P. christii* needs to be investigated (Kruskop & Lavrenchenko, 2000). Benda et al. (2011) recognize the Eritrean form as *P. balensis* and they tentatively assigned *Plecotus* specimens from Yemen as *Plecotus* cf. *balensis*. During a bat survey in central Ethiopia, we were able to trap several species, among them phenotypically *P. balensis*. The aim of our study was to verify the species identification with molecular methods and to verify their position within the *Plecotus* phylogeny.

Material and methods

Ethical statement

We obtained ethical approval from the Review Board of Addis Ababa University Institutes (IRB) for trapping bats and sacrificing and depositing a few individuals as voucher specimens in the Genetics Laboratory of the Department of Zoological Science at the University of Addis Ababa (Reference number: CNCSDO/181/14/2021).

Trapping

During a bat survey along the Ethiopian Rift Valley and adjacent highlands, we trapped bats at 55 sites in 12 regions (Fig. 1). We set up mist nets in and around roosting sites, such as caves, churches, and lodge sites. We used five mist nets, each 12 m long and 3 m wide. Mist netting was conducted between 7:00 pm and 9:00 pm for 7 to 13 nights per site. We tentatively identified the trapped bats in the field based on phenotype (Fig. 2) and morphological measurements. After measuring the bats, we immediately released them into their natural environment, except for some voucher specimens. From these specimens, we took tissue samples for a molecular analysis to verify phenotypic identification. We preserved the fresh tissue (liver, kidney, and heart) in 18 ml plastic tubes with 97% ethanol for further processing. The samples were shipped to the genetics lab of the German Primates Center for molecular species identification.



Figure 1. Bat trapping sites in 12 regions in central Ethiopia. *Plecotus balensis* was only trapped in regions 10-12. (Green colours indicate lower, brown colours higher altitude).



Figure 2. Pictures of the Ethiopian long-eared bat *Plecotus balensis* (a–f face views; g and h ventral views; i dorsal view; j lateral view). Voucher specimens have been deposited in the Genetics Laboratory of the Department of Zoological Science at the University of Addis Ababa, Ethiopia) (Photos by Ahmed Seid Ahmed)

Morphometrics

After capturing the bats, we placed them individually in cotton bags. We took morphometric measurements of the external morphology from those individuals that were sacrificed using rulers, ensuring measurements with adjustments made to the closest 0.1 cm. Body mass was taken with a Pesola spring scale.

DNA extraction and sequencing

We extracted DNA from tissue samples using the NucleoSpin Tissue Mini Kit (Macherey-Nagel) following the standard protocol of the supplier including the recommended sample digestion overnight at 56°C and 900 rpm (Thermomixer Comfort; Eppendorf). We amplified the complete mitochondrial cytochrome b gene (cytb, 1140 bp) with primers 5'-

AAAAAYCACCGTTGAYTTCAAC-3' and 5'-ATTACACTGGTCTTGTAAACCA-3' via polymerase chain reaction (PCR). The PCR reaction with a total volume of 30 μ L contained 3 μ L buffer (10× with 18 mM MgCl₂), 0.6 μ L dNTPs (10 mM each), 1 μ L of each primer (1 μ M), 0.3 μ L (1 U) FastStart HighFidelity Taq DNA polymerase (Roche), and 50 ng genomic DNA. Amplification was performed in a laboratory cycler (SensoQuest) with a pre-denaturation step at 94°C for 2 min, followed by 40 cycles at 94°C for 1 min, 56°C for 1 min, and 72°C for 1.5 min, and terminated with a final extension step at 72°C for 5 min. PCR products were size-separated on 2% agarose gels, excised from the gel, purified with the Monarch DNA Gel Extraction Kit (New England Biolabs), and sent to Eurofins Genomics for Sanger sequencing using both amplification primers. Obtained sequence electropherograms were checked and manually corrected with SOFTWARE (Logsdon et al., 2011). Newly generated sequences have been deposited in GenBank (Accession Numbers PP212955 - PP212962).

Molecular species identification

For the molecular species identification, we compared our newly generated cytb haplotypes with sequences deposited in GenBank using the nucleotide Basic Local Alignment Search Tool (BLAST; https://blast.ncbi.nlm.nih.gov/Blast.cgi) of the National Library of Medicine (NCBI). Among the sequences with the closest matches, we found 33 unique haplotypes of *P. balensis*. To explore the phylogenetic affinities of the newly detected *P. balensis* specimens, we downloaded orthologous sequences of *P. balensis* and other *Plecotus* species from GenBank. A drawback of the phylogenetic reconstruction was the fact that for most closely related species, the available data consisted of only partial cytb sequences (e.g., *P. balensis* 650 bp, *P. christii* 522 bp), which even did not overlap completely. Therefore, the length of the resulting alignment was only 492bp. As an outgroup, we used closely related species of the Vespertilioninae *Barbastella leucomelas*, *Glauconycteris variegate*, and *Scotoecus hirundo* (Table 1). The complete alignment consisted of 68 sequences.

Taxon	Group	Length	Id / haplo	Provenance	Lat	Long	AccNum	Source		
P. balensis	austriacus	1140	GU1 / 01	Guassa	10.29119	39.78599	8599 PP212955 thi			
P. balensis	austriacus	1140	ET59/03 Entoto 9.08922 38.76447 PP212956					this study		
P. balensis	austriacus	1140	MF37 / 04 Menagesha 8.96730 38.54892 PP212957							
P. balensis	austriacus	1140	MF36 / 04 Menagesha 8.96730 38.54892 PP212958					this study		
P. balensis	austriacus	1140	SF46 / 02 Sansusi 9.08155 38.69510 PP212959					this study		
P. balensis	austriacus	1140	SF47 / 03 Sansusi 9.08155 38.69510 PP21					this study		
P. balensis	austriacus	1140	SF48 / 03	PP212961	this study					
P. balensis	austriacus	1140	SF49 / 03	PP212962	this study					
P. balensis	austriacus	650		Bale Mts			MW166392	[1]		
P. balensis	austriacus	650		Bale Mts			MW166395	[1]		
P. balensis	austriacus	650		Bale Mts			MW166396	[1]		
P. balensis	austriacus	650		Bale Mts			MW166397	[1]		
P. balensis	austriacus	650		Bale Mts			MW166398	[1]		
P. balensis	austriacus	650		Bale Mts			MW166393	[1]		
P. balensis	austriacus	650		Bale Mts			MW166394	[1]		
P. balensis	austriacus	650		Bale Mts			MW166384	[1]		
P. balensis	austriacus	650		Bale Mts			MW166383	[1]		
P. balensis	austriacus	650		Bale Mts			MW166385	[1]		
P. balensis	austriacus	650		MW166386	[1]					
P. balensis	austriacus	650		MW166432	[1]					
P. balensis	austriacus	650		Mt Chilalo			MW166402	[1]		
P. balensis	austriacus	650		Guassa			MW166425	[1]		
P. balensis	austriacus	650		Guassa			MW166430	[1]		
P. balensis	austriacus	650		Guassa			MW166427	[1]		
P. balensis	austriacus	650		MW166426	[1]					
P. balensis	austriacus	650		MW166433	[1]					
P. balensis	austriacus	650		MW166417	[1]					
P. balensis	austriacus	650		MW166416	[1]					
P. balensis	austriacus	650		Mt Abune	MW166420	[1]				
P. balensis	austriacus	650		Mt Abune	MW166422	[1]				
P. balensis	austriacus	650		Mt Abune			MW166423	[1]		
P. balensis	austriacus	650		Mt Abune			MW166418	[1]		
P. balensis	austriacus	680		Mt Abune			AF413798	[2]		
P. balensis	austriacus	680		Mt Abune			AF413799	[2]		
P. balensis	austriacus	650		Simien Mts			MW166405	[1]		
P. balensis	austriacus	650		Simien Mts			MW166410	[1]		
P. balensis	austriacus	650		Simien Mts			MW166409	[1]		
P. balensis	austriacus	650		Simien Mts			MW166413	[1]		
P. balensis	austriacus	650		Simien Mts			MW166408	[1]		
P. balensis	austriacus	650		Simien Mts			MW166415	[1]		
P. balensis	austriacus	650		Simien Mts			MW166404	[1]		
P. balensis	austriacus	650		Simien Mts			MW166406	[1]		
P. christii	austriacus	522		Egypt, Sinai			EU743799	[3]		
P. christii	austriacus	522		Libya			EU743800	[3]		
P. christii	austriacus	522		Jordan			EU743801	[3]		
P. austriacus	austriacus	680		Spain			AF513788	[2]		

Table 1. Origin, sequence lengths (bp), and GenBank accession numbers of bat samples.

P. austriacus	austriacus	680	Portugal	AF513786	[2]
P. kolombatovici	austriacus	680	Turkey	AF513785	[2]
P. teneriffae	austriacus	680	Canary Is.	EU360701	[4]
P. auritus	auritus	1140	Denmark	MT410875	[5]
P. auritus	auritus	1140	Denmark	MN122881	[5]
P. auritus	auritus	1140	Switzerland	OQ885414	[6]
P. auritus	auritus	1140	Switzerland	OQ885415	[6]
P. auritus	auritus	1140	?	OY734038	[7]
P. macrobullaris	auritus	1140	Italy	KR134353	[8]
P. macrobullaris	auritus	1140	Turkey	KR134407	[8]
P. macrobullaris	auritus	1140	Syria	KR134406	[8]
P. turkmenicus	auritus	1140	Mongolia	MT583351	[9]
P. kozlovi	auritus	1140	Mongolia	MT583349	[9]
P. ognevi	auritus	1140	Mongolia	MT583350	[9]
P. (auritus) ognevi	auritus	1140	Korea	HM164052	[10]
P. homochrous	auritus	1140	Vietnam	MN160089	[11]
P. homochrous	auritus	1140	Vietnam	MN160086	[11]
P. sacrimontis	auritus	1140	Japan	LC036641	[12]
P. (auritus) sacrimontis	auritus	1140	Japan	AB085734	[13]
Barbastella leucomelas		1140	China	KU922958	[14]
Glauconycteris variegata		1140	Senegal	JX276108	[15]
Scotoecus hirundo		1140		JX276317	[15]

1 Razgour et al., 2021; 2 Juste et al., 2004; 3 Benda et al., 2008; 4 Garcia-Mudarra et al., 2009; 5 Margaryan 2020, unpubl.; 6 Ruedi et al., 2023; 7 2023, unpubl.; 8 Alberdi et al., 2015; 9 Kruskop et al., 2020; 10 Hwang & Kim 2016, unpubl.; 11 Fukui et al., 2020; 12 Kawai 2015, unpubl.; 13 Sakai et al., 2003; 14 Liu 2017, unpupl.; 15 Koubínová et al., 2013

In a second approach, we reconstructed a phylogeny based on the complete cytb (1140 bp). However, due to the limited number of full-length cytb sequences, the alignment comprised only 27 sequences with our *P. balensis* sequences as the only representatives of the *austriacus* group (see supplement). Maximum-likelihood trees were reconstructed with IQ-tree 2.2.0 (Minh et al., 2020) using the best-fit model as obtained by ModelFinder (Kalyaanamoorthy et al., 2017) in IQ-tree under the Bayesian Information Criterion (BIC) and 1000 ultrafast bootstrap replications (Hoang et al., 2018). Best-fit models for the alignments containing 492 bp and 1140 bp were TPM2+F+I+G4 and K3Pu+F+I+G4, respectively. Trees were visualized and edited with FigTree 1.4.0 (http://tree.bio.ed.ac.uk/software/figtree/).



Figure 3. Distribution of Plecotus balensis in the Ethiopian highlands. The taxa at the historical sites have been listed as *Plecotus auritus*, *P. austriacus*, or *P. christii* (Sordelli 1902; Senna 1905; Largen et al., 1974; Yalden et al., 1996). (1) Simien Mountains; (2) Mount Abune Yosef; (3) Guassa Community Conservation Area; (4) Menagesha, Entoto, Sansusi; (5) Chilallo Mountains; (6) Bale Mountains. (Green colours indicate lower, brown colours higher altitude).

Results

In three of the 12 trapping areas, we caught *P. balensis*. These areas were in the central highlands of Ethiopia at altitudes between 2500 m and 3300 m [Menagasha (region 10), Entoto/Sansusi (region 11), Guassa CCA (region 12)] (Fig. 3). The average body size of our ten measured individuals tends to be slightly larger than that of the holotype (Table 2).

ID	Site	Latitude	Longitude	Sex	FA	TB	FL	EL	TL	BL	TR	WS	BM	cytb
					mm	mm	mm	mm	mm	mm	mm	mm	g	
holo	Harenna	6.75000	39.73333	М	36.3	14.4	7.1/8.3	37	48	45	15.2			
ET59	Entoto	9.08922	38.76447	М	41.5	18	8.5	40	46	47	19	290	7.5	yes
MF36	Menagesha	8.96730	38.54892	F	40	17.5	9	39	51	49	19	285	8	yes
MF37	Menagesha	8.96730	38.54892	М	41	18	8	41	50	48.5	18.5	284	7.5	yes
MF39	Menagesha	8.96730	38.54892	М	40	18	8	40	47	51	19	284	7.7	no
SF46	Sansusi	9.08155	38.69510	М	41	19	9	37	41	53	18	282	8	yes
SF47	Sansusi	9.08155	38.69510	М	40	18.5	8.7	39	50	49	19	285	8.5	yes
SF48	Sansusi	9.08071	38.69455	F	39.5	17.5	8	41	48.5	48	18	287	7.5	yes
SF49	Sansusi	9.08071	38.69455	М	41	18.4	8.3	41.5	49	50	19	284	7.9	yes
GU1	Guassa	10.29119	39.78599	М	40	19	9	38	42	52	18.5	283	7.5	yes
GU3	Guassa	10.17490	39.47450	F	38	17.5	7.5	36.5	42	46	18	279	7	no
Mean					40.2	18.1	8.4	39.3	46.7	49.4	18.6	284.3	7.7	
SD					1.00	0.57	0.52	1.72	3.74	2.19	0.14	2.76	0.39	

Table 2. Geographic coordinates (decimal degree) of origin and morphological measurements of ten *Plecotus balensis* individuals and the holotype (Kruskop and Lavrenchenko, 2000).

FA = forearm length; TB = tibia length; FL = foot length; EL = ear length; TL = tail length: BL = body length; TR = tragus; WS = wingspan; BM = body mass.

Genetic taxon identification

We sequenced the complete cytb gene from eight individuals from which we retrieved four haplotypes. Using BLAST search, the greatest matches (99.5 and 99.6% identity, query cover 57% and 59%, accession length 650 bp and 680 bp, respectively) were found with the partial cytb sequences of *P. balensis* provided by Razgour et al. (2021) and Juste et al. (2004). The phylogenetic tree based on 492 bp (Fig. 4) confirms a well-supported monophyly for *P. balensis* and suggests *P. christii* as the sister taxon. As expected, *P. balensis* falls in the *austriacus* species group. Due to the short sequence lengths, our eight *P. balensis* sequences comprised only two haplotypes of which one (GU1) from Guassa was identical to haplotype Guassa02 of Razgour et al. (2021), whereas the second haplotype found at Sansusi, Entoto (region 11) and Menagesha (region 10) was distinct. The mitochondrial intra-specific phylogenetic relationships of *P. balensis* seem to be geographically ordered with a division into a southern, central, and northern deme. However, the statistical support for most of the nodes in the tree is weak.



Figure 4. Phylogenetic relationships of the newly generated *Plecotus balensis* haplotypes (marked red) based on partial cytb sequences (492 bp). Node labels refer to ML bootstrap support values. Orange colours indicate northern samples, blue colours indicate central samples and green colours indicate southern samples. (CHE = Switzerland; DNK = Denmark; EGY = Egypt; ETH = Ethiopia; ESP = Spain; ITA = Italia; JOR = Jordan; JPN = Japan; KOR = South Korea; LBY = Libya; MNG = Mongolia; PRT = Portugal; SEN = Senegal; SYR = Syria; TUR = Turkey)

For the complete cytb sequences (1140 bp) we retrieved four haplotypes which form together a well-supported clade within the genus *Plecotus* (Fig. 5). In this phylogeny, the major division was between *P. balensis* as a representative of the *austriacus* group and taxa of the *auritus* group. Within the *auritus* group, we found four similarly well-supported clades. One consisted

of *Plecotus* taxa from central and northern Europe (*P. auritus*), a second from the Mediterranean and Near East region (*P. macrobullaris*), a third (*P. sacrimontis*), and fourth (*P. kozlovi*, *P. ognevi*) from East Asia. However, the deeper splits and the relationships of *P. turkmenicus* and *P. homochrous* within the clade are not well supported.



Figure 5. Phylogenetic relationships of *Plecotus* spp. mtDNA lineages based on complete cytb sequences (1140 bp). *Plecotus balensis* (marked blue) forms a strongly supported clade within the genus *Plecotus* but the relationships among the Plecotus clades are only weakly supported. Node labels refer to ML bootstrap support values. (CHE =Switzerland; DNK = Denmark; ETH = Ethiopia; ITA = Italia; JPN = Japan; KOR = South Korea; MNG = Mongolia; SEN = Senegal; SYR = Syria; TUR = Turkey; VNM = Vietnam)

Discussion

Our results correspond well with the previous studies on long-eared bats from Ethiopia and other regions. We confirmed the presence of *P. balensis* in areas 25 km west and 10 km north of the centre of Ethiopia's capital, Addis Abeba. This area is about 180 km and 200 km, respectively, southwest of the closest known site, Guassa CCA (Razgour et al. 2021). Rüppel (1842) collected *Plecotus auritus* (most likely *P. balensis*) in the historical "Shoa" (Shewa) region and our new *Plecotus* sites close to Addis Abeba are within this region. It would be interesting to obtain genetic information from the Rüppell specimen and the Bale Mountains specimen deposited at the museum in Liverpool (Yalden et al. 1996). In general, our samples add new mitochondrial haplotypes to the genetic diversity of *P. balensis* and support a

geographic pattern of haplotype distribution. It is most likely that the Ethiopian population divides into a southern, central, and northern deme. This pattern reminds us of the distribution of the genetic diversity of another Ethiopian high-altitude species, *Theropithecus gelada* (Zinner et al., 2018; Trede et al., 2020). However, since only relatively few mountain areas have been surveyed for the presence of *P. balensis*, this pattern might change and turn into a clinal genetic distribution. At least the ecological niche model by Razgour et al. (2021) suggests suitable habitats in several other mountainous areas of Ethiopia, e.g., Gojjam and Ahmar Mountains, and a much wider distribution of the species during the last glacial maximum. It remains to be investigated whether the distribution range of *P. balensis* extends further north into Eritrea and whether an area of sympatry of the two species exists.

Our *Plecotus* specimens tend to be slightly larger than the holotype, which comes from the Bale Mountains. Whether the *Plecotus* demes actually differ in body size needs to be further explored. On the other side, we took the morphometric measurement with a ruler, which is most likely more error-prone than other methods. Although our current analysis is mainly based on a relatively short part of the cytb gene, it nevertheless strongly supports the identification of our samples as *P. balensis*. Our phylogenetic reconstruction indicates a sister relationship with *P. christii*, which is not surprising, given that it is geographically the closest species. However, here we need to be cautious, because of the limited genetic information. The Ethiopian long-eared bat is currently classified as Data Deficient by the IUCN (Lavrenchenko et al. 2019). A denser sampling of *Plecosus* in Ethiopia would improve the information on the status of this and other species of the genus. Additionally, sampling in Eritrea, northeastern Sudan, and the western mountains of the Arabian Peninsula and the inclusion of additional genetic markers would be important, and essential to answer open questions about the phylogeny of long-eared bats in the region around the Red Sea.

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